

Enzymatic resolution of bicyclo[n.1.0]alkan-1-ols derivatives: Preparation of optically active α -substituted α -methylcycloalkanones.

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Abstract : Optically active α -methyl α -substituted cycloalkanones are prepared by a chemoenzymatic sequence which involves a Lipozyme®-catalyzed transesterification of 1-(chloroacetoxy)bicyclo[n.1.0]alkanes and ring opening of these cyclopropanol derivatives. © 1998 Elsevier Science Ltd. All rights reserved.

Optically active α,α -disubstituted cycloalkanones are useful intermediates for the synthesis of natural and/or biologically active compounds bearing a quaternary chiral carbon atom.^{1,2,3} Such optically active cycloalkanones^{4,5,6} could be prepared by alkylation of ketonic compounds in the presence of a chiral catalyst⁷ or using a chiral base,⁸ by alkylation of ketone enolate derivatives with alkylating agents bearing a chiral group,⁹ by reaction of metallated¹⁰ or unmetallated¹¹ chiral enamines or imines derivatives with alkylating agents, by addition of allyl derivatives to β -dicarbonyl compounds in the presence of palladium complexes bearing chiral ligands, by asymmetric Claisen rearrangement¹² and by enzymatic reduction of 2,2-disubstituted 1,3-diketones.^{13,14} In fact most of these methods gave access to cycloalkanones bearing on the quaternary carbon atom an activating group (ester, ketone, nitrile, phenyl, ...) and/or an alkyl chain with an electron withdrawing group in β -position. The former groups were introduced in order to allow the alkylation of a α -monosubstituted cycloalkanone and/or to obtain a regioselective addition and the latter groups came from the addition of electron poor alkenic reactants. When focus was put on α -substituted α -methylcycloalkanones, the synthesis using these methods should necessitate various reactions including protecting and deprotecting steps in order to obtain the substituents in the proper oxidation state and with the desired chain length.

Recently it has been reported that lipase-catalysed transesterification of bicyclo[4.1.0]heptan-1-yl chloroacetates allows the preparation of optically active bicyclo[4.1.0]heptan-1-ol derivatives which can be transformed into enantiomerically enriched 2-methylcyclohexanones by the well known cleavage of cyclopropanols in basic medium. When an alkyl substituent was present on the bicyclic junction carbon atom α to the hydroxyl or to the chloroacetoxy group, α -substituted α -methylcyclohexanones were obtained and no epimerisation was observed during the cyclopropanic bond cleavage.¹⁵

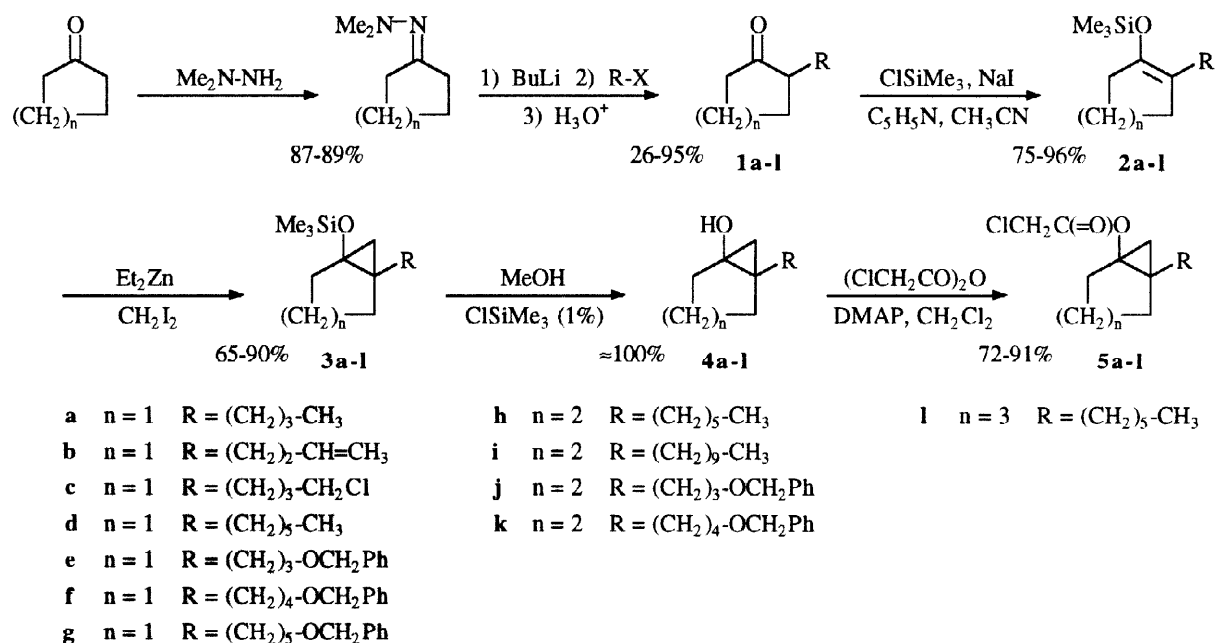
This approach to α -substituted α -methylcycloalkanones seems interesting because the substituent R introduced during the synthesis of the bicycloalkanyl chloroacetate precursor could present the desired

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functionalities and the appropriate chain length to avoid further tedious chemical transformations on this moiety.

Herein is reported an examination of the scope of this synthetic method with respect to the size of the bicycloalkane core and to the chain length and the functionality of the substituent.

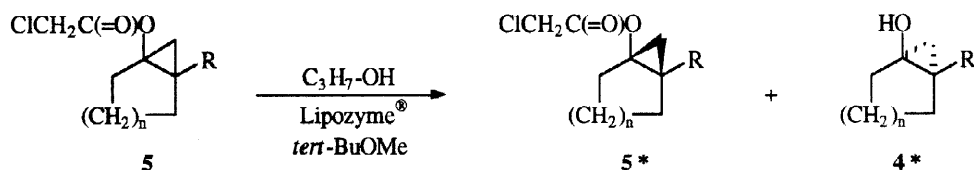
The bicyclo[n.1.0]alkan-1-yl chloroacetates **5a-l** were prepared from 2-substituted cycloalkanones **1a-l** as shown in Scheme 1.



Scheme 1. Preparation of 1-(chloroacetoxy)bicyclo[n.1.0]alkanes **5a-l**

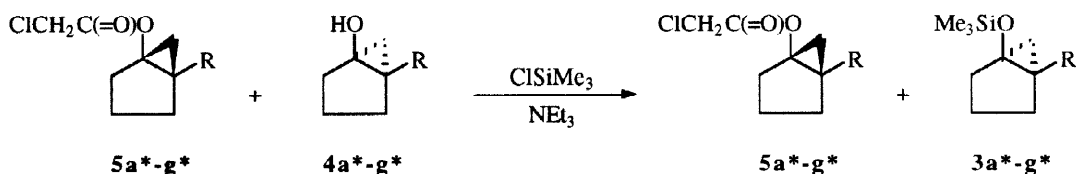
The α -substituted cycloalkanones **1a-l** were obtained by addition of various alkylating agents to the lithium derivatives¹⁶ of cyclopentanone, cyclohexanone and cycloheptanone dimethylhydrazone.¹⁷ Hydrolysis of the alkylated hydrazone to obtain the corresponding α -substituted cycloalkanone was made using an aqueous solution of potassium monophosphate and phosphorus acid.¹⁸ The α -substituted silyl enol ethers **2a-l**, synthesized by reaction of the 2-substituted cycloalkanones **1a-l** with chlorotrimethylsilane, sodium iodide and pyridine in acetonitrile,^{19,20} were cyclopropanated by a diethylzinc-methylene iodide mixture²¹ to give the corresponding 1-(trimethylsilyloxy)bicyclo[n.1.0]alkanes **3a-l**. The beneficial influence of oxygen in the cyclopropanating reaction should be emphasized.²² When the reaction was run under argon, low reaction rates and low yields of cyclopropanated products were generally observed and in some cases no cyclopropanation has occurred.^{23,24} Treatment of the trimethylsilyl ethers **3a-l** by methanol acidified by addition of $ClSiMe_3$ (1 mole %) gave cleanly the bicycloalkan-1-ols **4a-l** which are isolated²⁵ after addition of NEt_3 (1 mole %) ²⁶ and esterified with chloroacetic anhydride in the presence of 4-dimethylaminopyridine. Chloroacetates **5a-l**²⁷ were characterized by their 1H and ^{13}C NMR, IR and mass spectra.

Transesterifications of chloroacetates **5a-l** by 1-propanol were run in *tert*-butyl methyl ether at 36°C in the presence of Lipozyme®²⁸ which was found to be an appropriate lipase to catalyze the transesterification of bicyclo[4.1.0]heptan-1-yl chloroacetates.¹⁵ Generally, reactions were stopped after about 50% conversion by removing the enzyme by filtration.



In the case of the bicycloheptanes and the bicyclooctane the separation of the unreacted chloroacetate **5*** ($n = 2, 3$) and the bicycloalkanol **4*** ($n = 2, 3$) was made by chromatography on neutral alumina (Activity III). (On silica gel, transformation of the bicycloalkanol into 2-methylcycloalkanone occurs).

Generally, bicyclohexanols **4*** ($n = 1$) are not recovered after chromatography under usual conditions (on silica gel, alumina or Florisil®) because they are air-sensitive.²⁹ After transesterification of the bicyclohexanes **5a-g** the mixtures of optically active products were treated under argon with chlorotrimethylsilane in the presence of triethylamine in ether, then the silyloxybicyclohexane **3*** was easily separated from the chloroacetate **5*** by chromatography on Florisil® (silyloxycyclopropanes were partly decomposed on silica gel or alumina).



In Table I are collected our results and those concerning other (chloroacetoxy)bicycloheptanes previously reported.¹⁵ Generally no transesterification of the isomeric (chloroacetoxy)bicycloalkanes synthesized from the minor less substituted silyl enol ethers were observed. The remaining optically active chloroacetates **5*** were always isolated mixed with their regioisomers and the bicycloalkanols **4*** or the silyl ethers **3*** were obtained in pure form. Enantiomeric purities of the remaining chloroacetates **5*** and those of optically active bicycloalkanols **4*** or the corresponding trimethylsilylethers **3*** were determined after transformation into 2-substituted 2-methylcycloalkanones **6** (vide infra).

The reaction rate of the transesterification catalyzed by the lipase of *Mucor miehei* was dependent on the starting chloroacetate ring size. In the conditions used in this work (see experimental part) about 50% conversion was reached after 8 hours for the cyclopentanic and the cyclohexanic compounds **5a-5k** and 36% conversion only was observed after 48 hours in the case of the cycloheptanic compound **5l**.³¹

In the cases studied, moderate to excellent ee's were found for the remaining chloroacetates **5*** and for the bicycloalkanols **4*** or the silylethers **3***. Enantioselectivity of the reaction appears generally better for the cyclopentanic compounds **5a-5g** (enantiomeric ratio E between 40 and 420) than for the cyclohexanic compounds **5h-5k** (E between 14 and 20). For the much slower reacting cycloheptanic chloroacetate **5l** the enantiomeric ratio (E = 19) was closed to that calculated for the cyclohexanic homolog **5h** (E = 18) which bears the same substituent.

For a same size of the bicycloalkane the enantioselectivity of the reaction is chain length dependent. In the case of the cyclohexanic compounds it appears that the enantiomeric ratio E increases from the unsubstituted chloroacetate (**5**, $n = 2$, R = H, E = 22) to the compound substituted with a butyl chain (**5**, $n = 2$, R = C₄H₉, E = 38) then the E value decreases when the substituent is longer (**5h** : E = 18; **5i** : E = 14). The same decrease of the E value was observed with cyclopentanic compounds when the alkyl chain increases from butyl (**5a** : E = 420) to

Table I. Transesterification of bicyclo[n.1.0]alkan-1-yl chloroacetates **5** by 1-propanol in the presence of Lipozyme® (Lipase from *Mucor miehei*)^a

Substrate 5			Remaining Chloroacetate 5*			Product (see text)				
n	R	c ^d	Yield (%)	[α] _D ²⁰ ^e	ee _s (%)	4* (P = H) or 3* (P = SiMe ₃)				E ^d
						P	Yield (%)	[α] _D ²⁰ ^e	ee _p (%)	
1	(CH ₂) ₃ -CH ₃ 5a	0.50	47	+14.2° (0.5)	97	SiMe ₃	42	+0.7° (0.5)	98	420
1	(CH ₂) ₂ -CH=CH ₂ 5b	0.53	46	+11.6° (1)	97	SiMe ₃	38	+1.3° (1)	86	55
1	(CH ₂) ₃ -CH ₂ Cl 5c	0.49	37	+9.3° (2)	93	SiMe ₃	29	+1.8° (1)	94	110
1	(CH ₂) ₅ -CH ₃ 5d	0.46	39	+7.7° (1)	81	SiMe ₃	41	+0.9° (1)	97	165
1	(CH ₂) ₃ -OCH ₂ Ph 5e	0.51	36	+6.4° (1)	91	SiMe ₃	42	+2.3° (1.5)	87	46
1	(CH ₂) ₄ -OCH ₂ Ph 5f	0.53	50	+10.2° (2)	95	SiMe ₃	49	+2.1° (1)	85	45
1	(CH ₂) ₅ -OCH ₂ Ph 5g	0.51	48	+6.7° (1)	90	SiMe ₃	40	+2.0° (1)	86	40
2 ^c	H	0.51		+10.4° (1)	84	H		+5.7° (1)	79	22
2 ^c	CH ₃	0.57		+30.2° (1)	95	H		-18.5° (1)	72	22
2 ^c	C ₂ H ₅	0.53		+18.5° (1)	91	H		-10.5° (1)	80	28
2 ^c	(CH ₂) ₃ -CH ₃	0.51		+18.5° (1)	90	H		-11.6° (1)	86	38
2 ^c	(CH ₂) ₅ -CH ₃ 5h	0.56		+13.0° (1)	86	H		-8.0° (1)	66	13(18) ^f
2	(CH ₂) ₉ -CH ₃ 5i	0.54	49	+12.6° (1.3)	86	H	43	-6.4° (0.9)	74	14
2	(CH ₂) ₃ -OCH ₂ Ph 5j	0.54	52	+12.5° (1.5)	88	H	42	-11.1° (1)	75	20
2	(CH ₂) ₄ -OCH ₂ Ph 5k	0.55	54	+14.2° (1.4)	85	H	41	-9.1° (1.3)	70	15
3 ^b	(CH ₂) ₅ -CH ₃ 5l	0.38	76	-1.3 (0.9)	51	H	28	-3.7 (0.8)	84	19

a) Unless otherwise noted the reaction time was about 8 hours.

b) 48 hours reaction time.

c) From reference 15.

d) conversion (c) and E calculated from ee_s and ee_p according to ref. 30.

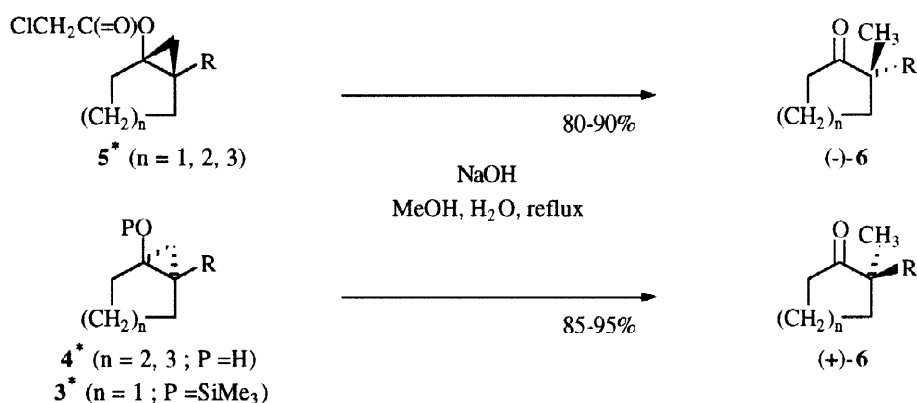
e) In parenthesis is indicated the concentration in THF.

f) Using the conditions of this work.

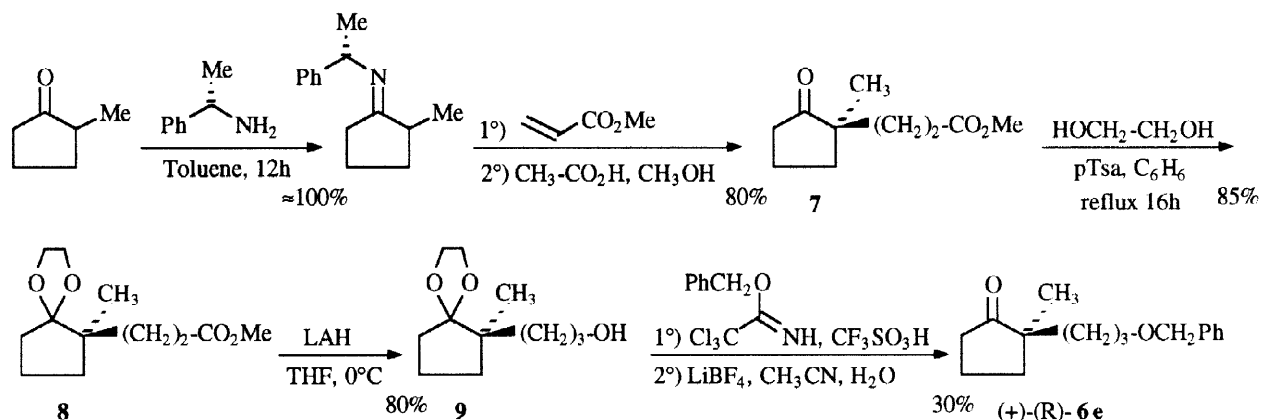
hexyl (**5d** : E = 165) and with the cyclopentanic and the cyclohexanic chloroacetates when the ω-benzyloxyalkyl chain length increases (compare E values calculated for the reactions of **5e**, **5f** and **5g** in one part and those of **5j** and **5k** in another part). It is also noteworthy that the presence of a carbon-carbon double bond or an heteroatom at the end of a butyl chain decreases the enantioselectivity of the reaction (compare the E values calculated for the reactions of **5b** and **5c** to that of **5a**).

Treatment of optically active chloroacetates **5***, bicycloalkanols **4*** and trimethylsilyl ethers **3*** with 2 equivalents of sodium hydroxide in refluxing hydromethanolic solutions led to the enantiomerically enriched 2-substituted 2-methylcycloalkanones **6a-6l** (yields ≥ 80%). At this point the two isomeric α,α'-disubstituted cycloalkanones which were always present in the reaction mixture obtained from a chloroacetate were generally separated from the cycloalkanone **6** during purification by silica gel column chromatography. The ee's of the ketones **6e**, **6g** and **6j** were measured by ¹H NMR spectroscopy in the presence of Eu(hfc)₃. For all the other

ketones ee's were determined by gas chromatography on a chiral column (Lipodex E). From the chloroacetoxybicycloalkanes **5*** levorotatory cycloalkanones were obtained and dextrorotatory compounds were isolated from the bicycloalkanol **4*** or the trimethylsilyl ethers **3***.



A sample of the cyclopentanone **6e** enriched in the R-configured isomer was synthesized starting from the known (R)-ketoester **7**³ which was easily obtained by reaction of methyl acrylate with the imine resulting from the reaction of 2-methylcyclopentanone with (S)-1-phenylethylamine. After formation of a dioxolane protecting group, the ester function was reduced,³² then the hydroxyl group was transformed into a benzyloxy group³³ and the ketone was deprotected (Scheme 2).

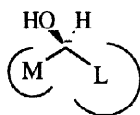


Scheme 2. Synthesis of cyclopentanone **6e** of known configuration

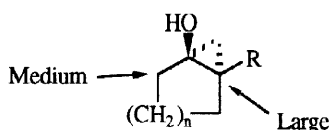
This (R)-cyclopentanone **6e** shows the same positive specific rotation in THF and the same retention time in gas chromatography on a Lipodex E column than the main isomer isolated after treatment in basic medium of the trimethylsilyl ether **3e*** which was obtained from the bicyclohexanol formed in the Lipzyme®-catalyzed transesterification of **5e**. So the transesterification of **5e** has occurred mainly on the (1S, 5R) enantiomer. The same enantioselectivity was observed in the Lipzyme®-catalyzed transesterification of the chloroacetoxybicyclo[4.1.0]heptanes **5** with $\text{R} = \text{H}$ or C_4H_9 ¹⁵ and was postulated in all the other cases. The homogeneity of the specific rotation sign of silyloxybicycloalkanes **3a*-3g*** (dextrorotatory) in one part, of bicycloalkanol **4h*-4l*** (levorotatory) in another part and of all the cycloalkanones **6** obtained from the more reactive chloroacetoxybicycloalkane enantiomers were in agreement with such an assumption.

An empirical rule predicts which enantiomer of secondary alcohols reacts faster with the lipase from *Mucor miehei*.³⁴ When the hydroxyl group points up and forward out of the plane defined by the chiral carbon

center, the large and the medium substituents, the favored enantiomer bears the large substituent on the right.



Generally in the transesterification of secondary alcohol esters it is the enantiomer with the same configuration that reacts faster. To our knowledge this empirical rule was not extended to tertiary alcohols or their derivatives because few examples of enzyme-catalyzed reactions of such compounds were reported in the literature.³⁵ In the case of the bicyclo[n.1.0]alkan-1-ol derivatives it appears that the fast reacting enantiomer presents the same characteristics than above, if we consider that the polysubstituted C_{n+2} cyclopropanic carbon atom is the larger substituent and the C₂ methylene group the medium substituent.



In conclusion this method allows the selective preparation of the two enantiomers of α -methylcycloalkanones of various size α -substituted by various substituents.

Experimental Section

General Methods. ¹H and ¹³C NMR spectra were recorded on a Bruker AC-200 or on a AC-250 instrument. Chemical shifts are expressed relative to tetramethylsilane. Abbreviations are as follows : s, singlet ; d, doublet ; dd, doublet of doublet ; t, triplet ; tt, triplet of triplet ; m, multiplet. Infrared spectra were recorded on a Perkin-Elmer 682 spectrometer. Mass spectra were determined on a GC-Mass spectrometer Nermag R10-10 (capillary column: CPSIL 5, 25 m) at an ionizing voltage of 70 eV. Column chromatography was carried out with 70-230 mesh silica gel, 100-200 mesh neutral alumina or 100-200 mesh Florisil. TLC was performed on 0.25-mm silica gel (Merk 60 F254). Dry solvents were obtained as follows : diethyl ether was distilled over LiAlH₄, THF was distilled over sodium-benzophenone radical-anion and pentane was distilled over P₂O₅. Triethylamine and pyridine were purified by distillation over CaH₂ and chlorotrimethylsilane by distillation over quinoline under argon. Other reagents were distilled before use. The dimethylhydrazones were prepared according to a literature procedure¹⁷ and the ω -benzyloxy-1-bromoalkanes were synthesized (1°- MeSO₂Cl, NEt₃, CH₂Cl₂. 2°- LiBr, acetone)³⁶ from ω -benzyloxy-1-alkanols.³⁷ The sequence to prepare the chloroacetate **5h** and the spectroscopic properties of the compound **5h** and the intermediates were previously reported.¹⁵ A 25 meters capillary column Lipodex E was used for the ee's measurements (flow carrier: helium).

Preparation of 2-butylcyclopentanone 1a. Representative Procedure. To a solution of 1.26 g of cyclopentanone dimethylhydrazone (10 mmol) in 20 mL of dry THF under argon was added 6.56 mL of a 1.6 M solution of n-BuLi (10.5 mmol) in hexane. After 1h at -30°C the mixture was allowed to warm to 0°C and 1.07 mL of 1-bromobutane (10 mmol) was added. After 2h at 25°C, 1.4 g of KH₂PO₄ (10 mmol) and 1.65 g of H₃PO₃ (20 mmol) were added, followed by 10 mL of water. This mixture was left overnight at 25°C. Water was added and the reaction mixture was extracted several times using ethyl ether. The combined organic phases were dried (Na₂SO₄) and the solvents were evaporated. The product was purified by silica gel column chromatography (pentane/ethyl ether : 80/20) to give 1.12 g (82%) of 2-butylcyclopentanone.³⁸

Cycloalkanones **1b - 1l** were prepared following the same procedure.

2-(But-3-ényl)cyclopentanone 1b: From cyclopentanone dimethylhydrazone and 4-bromo-1-butene: 1.04 g,

75% .³⁹

2-(4-Chlorobutyl)cyclopentanone 1c: From cyclopentanone dimethylhydrazone and 1-chloro-4-iodobutane: 0.454 g, 26%; IR (neat) 2940, 2860, 1785, 1450 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 3.55 (t, $J = 6.6$ Hz, 2H), 2.40–2.21 (m, 3H), 2.09–1.93 (m, 2H), 1.68–1.49 (m, 4H), 1.38–1.19 (m, 4H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 219.9, 48.3, 44.2, 37.4, 31.9, 28.9, 28.3, 24.2, 20.1; EI-MS (m/z) 176 (M^+ , 0.8), 174 (M^+ , 1.7), 97(5), 84 (100), 83 (27), 55 (27).

2-Hexylcyclopentanone 1d: From cyclopentanone dimethylhydrazone and 1-bromohexane: 1.38 g, 82%.⁴⁰

2-(3-Benzyloxypropyl)cyclopentanone 1e: From cyclopentanone dimethylhydrazone and 3-benzyloxy-1-bromopropane: 2.10 g, 91%; IR (neat) 2940, 2860, 1740, 1460, 1100 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 7.40–7.20 (m, 5H), 4.50 (s, 2H), 3.48 (t, $J = 6.3$ Hz, 2H), 2.40–1.90 (m, 5H), 1.90–1.20 (m, 6H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 221.9, 138.4, 128.2, 127.5, 127.4, 72.7, 70.1, 48.8, 37.9, 29.4, 27.6, 26.3, 20.6; EI-MS (m/z) 232 (M^+ , 2.3), 125 (4), 107 (6), 97 (31), 91 (100), 84 (17), 77 (4), 55 (10).

2-(4-Benzyloxybutyl)cyclopentanone 1f: From cyclopentanone dimethylhydrazone and 4-benzyloxy-1-bromobutane: 2.10 g, 87%; IR (neat) 2940, 2860, 1740, 1460, 1100 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 7.40–7.20 (m, 5H), 4.50 (s, 2H), 3.48 (t, $J = 6.4$ Hz, 2H), 2.40–1.90 (m, 5H), 1.90–1.20 (m, 8H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 220.6, 138.2, 127.9, 127.1, 127.0, 72.4, 69.7, 48.6, 37.6, 29.3, 29.1, 29.05, 23.8, 20.3; EI-MS (m/z) 246 (M^+ , 3), 139 (2), 111 (3), 107 (5), 97 (45), 91 (100), 84 (14), 77 (4), 55 (12).

2-(5-Benzyloxypentyl)cyclopentanone 1g: From cyclopentanone dimethylhydrazone and 5-benzyloxy-1-bromopentane: 1.92 g, 74%; IR (neat) 2930, 2860, 1740, 1455, 1100 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 7.42–7.29 (m, 5H), 4.50 (s, 2H), 3.48 (t, $J = 6.4$ Hz, 2H), 2.36–1.88 (m, 5H), 1.87–1.57 (m, 4H), 1.57–1.17 (m, 6H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 220.0, 128.2, 127.6, 126.8, 126.7, 72.1, 69.6, 48.3, 37.4, 29.0, 28.9, 26.7, 25.6, 20.1, 19.9; EI-MS (m/z) 260 (M^+ , 4), 107 (14), 97 (23), 91 (100), 84 (19), 83 (6), 77 (4), 55 (9).

2-Decylcyclohexanone 1i: From cyclohexanone dimethylhydrazone and 1-bromodecane: 2.14 g, 90%; IR (neat) 2930, 2860, 1715, 1455 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 2.45–2.18 (m, 3H), 2.18–1.90 (m, 2H), 1.90–1.50 (m, 4H), 1.45–1.05 (m, 18H), 0.85 (t, $J = 6.4$ Hz, 3H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 213.3, 50.6, 41.8, 33.7, 31.8, 29.7, 29.5, 29.4, 29.3, 29.2, 27.9, 27.1, 24.7, 22.5, 22.2, 13.9; EI-MS (m/z) 238 (M^+ , 7), 195 (3), 99 (79), 98 (100), 97 (25), 85 (4), 71 (6), 57 (11), 55 (22), 43 (20).

2-(3-Benzyloxypropyl)cyclohexanone 1j: From cyclohexanone dimethylhydrazone and 3-benzyloxy-1-bromopropane: 2.21 g, 90%; IR (neat) 2940, 2860, 1715, 1455, 1100 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 7.40–7.20 (m, 5H), 4.50 (s, 2H), 3.48 (t, $J = 6.5$ Hz, 2H), 2.49–2.20 (m, 3H), 2.20–1.95 (m, 2H), 1.95–1.75 (m, 2H), 1.75–1.50 (m, 4H), 1.50–1.22 (m, 2H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 213.0, 138.4, 128.1, 127.4, 127.3, 72.6, 70.2, 50.3, 41.8, 33.8, 27.9, 25.9, 24.7; EI-MS (m/z) 246 (M^+ , 5), 155 (14), 139 (19), 111 (27), 107 (5), 98 (17), 97 (11), 91 (100), 77 (10), 55 (46).

2-(4-Benzyloxybutyl)cyclohexanone 1k: From cyclohexanone dimethylhydrazone and 4-benzyloxy-1-bromobutane: 2.47 g, 95%.⁴¹

2-Hexylcycloheptanone 1l: From cycloheptanone dimethylhydrazone and 1-bromohexane: 1.63 g, 83%.⁴²

2-Substituted 1-(trimethylsilyloxy)cycloalkenes **2a–2l** were prepared from the 2-substituted cycloalkanones **1a–1l** using chlorotrimethylsilane, sodium iodide and pyridine in acetonitrile.¹⁹ In some cases the product has been purified by distillation, in the other cases the crude product was used in the next step.

2-Butyl-1-(trimethylsilyloxy)cyclopentene 2a: 1.74 g, 82%; IR (neat) 2960, 2840, 1690, 1455, 1350, 1260, 950, 750 cm^{-1} ; ^1H NMR (250 MHz, C_6D_6) δ 2.40–2.15 (m, 6H), 1.82–1.65 (m, 2H), 1.57–1.22 (m, 4H), 1.03–0.95 (m, 3H), 0.16 (s, 9H); EI-MS (m/z) 212 (M^+ , 5), 195 (6), 183 (61), 155 (34), 73 (100).

2-(3-Butenyl)-1-(trimethylsilyloxy)cyclopentene 2b: 1.68 g, 80%; IR (neat) 2960, 2840, 1690, 1650, 1455, 1350, 1260, 950, 750 cm^{-1} ; ^1H NMR (250 MHz, C_6D_6) δ 5.98–5.75 (m, 1H), 5.19–4.93 (m, 2H), 2.38–2.06

(m, 8H), 1.93–1.58 (m, 2H), 0.15 (s, 9H); EI-MS (*m/z*) 210 (*M*⁺, 11), 169 (6), 155 (33), 73 (100).

2-(4-Chlorobutyl)-1-(trimethylsilyloxy)cyclopentene 2c: 1.85 g, 75%; IR (neat) 2960, 2840, 1690, 1455, 1350, 1260, 950, 750 cm⁻¹; ¹H NMR (200 MHz, C₆D₆) δ 3.20 (t, *J* = 6.6 Hz, 2H), 2.36–1.98 (m, 6H), 1.80–1.64 (m, 2H), 1.64–1.28 (m, 4H), 0.15 (s, 9H); EI-MS (*m/z*) 248 (*M*⁺, 3), 246 (*M*⁺, 8), 169 (60), 73 (100).

2-Hexyl-1-(trimethylsilyloxy)cyclopentene 2d: 2.09 g, 87% .⁴⁰

2-(3-Benzoyloxypropyl)-1-(trimethylsilyloxy)cyclopentene 2e: 2.52 g, 83%; bp 130°C/0.04 mmHg; IR (neat) 2960, 2840, 1685, 1460, 1360, 1255, 1100, 950, 750 cm⁻¹; ¹H NMR (200 MHz, C₆D₆) δ 7.40–7.04 (m, 5H), 4.37 (s, 2H), 3.40 (t, *J* = 6.4 Hz, 2H), 2.40–2.14 (m, 6H), 1.91–1.52 (m, 4H), 0.16 (s, 9H); EI-MS (*m/z*) 304 (*M*⁺, 2), 169 (5), 107 (2), 91 (49), 89 (2), 73 (100).

2-(4-Benzoyloxybutyl)-1-(trimethylsilyloxy)cyclopentene 2f: 2.45 g, 77%; bp 145°C/0.04 mmHg; IR (neat) 2940, 2860, 1690, 1460, 1360, 1255, 1100, 950, 750 cm⁻¹; ¹H NMR (200 MHz, C₆D₆) δ 7.37–7.05 (m, 5H), 4.37 (s, 2H), 3.39 (t, *J* = 6.1 Hz, 2H), 2.45–2.05 (m, 6H), 1.82–1.46 (m, 6H), 0.16 (s, 9H); EI-MS (*m/z*) 318 (*M*⁺, 7), 211 (2), 183 (4), 107 (2), 91 (74), 89 (2), 77 (5), 73 (100).

2-(5-Benzoyloxypropyl)-1-(trimethylsilyloxy)cyclopentene 2g: 2.66 g, 80%; IR (neat) 2940, 2860, 1680, 1455, 1360, 1250, 1100, 950, 750 cm⁻¹; ¹H NMR (200 MHz, C₆D₆) δ 7.42–7.04 (m, 5H), 4.35 (s, 2H), 3.34 (t, *J* = 6.4 Hz, 2H), 2.42–2.11 (m, 6H), 1.85–1.58 (m, 4H), 1.52–1.39 (m, 4H), 0.16 (s, 9H); EI-MS (*m/z*) 332 (*M*⁺, 8), 225 (4), 107 (2), 91 (82), 89 (3), 77 (5), 73 (100).

2-Decyl-1-(trimethylsilyloxy)cyclohexene 2i: 2.33 g, 75%; bp 140°C/0.06 mmHg; IR (neat) 2940, 2865, 1680, 1450, 1355, 950, 750 cm⁻¹; ¹H NMR (200 MHz, C₆D₆) δ 2.27 (t, *J* = 7.3 Hz, 2H), 2.12–1.94 (m, 4H), 1.65–1.18 (m, 20H), 0.92 (t, *J* = 6.3 Hz, 3H), 0.16 (s, 9H); EI-MS (*m/z*) 310 (*M*⁺, 18), 295 (4), 267 (2), 197 (1), 183 (100), 169 (13), 75 (17), 73 (72).

2-(3-Benzoyloxypropyl)-1-(trimethylsilyloxy)cyclohexene 2j: 2.73 g, 88%; bp 142°C/0.06 mmHg; IR (neat) 2940, 2860, 1680, 1455, 1360, 1255, 1100, 950, 750 cm⁻¹; ¹H NMR (250 MHz, C₆D₆) δ 7.45–7.02 (m, 5H), 4.41 (s, 2H), 3.45 (t, *J* = 6.5 Hz, 2H), 2.32 (t, *J* = 6.5 Hz, 2H), 2.07–1.92 (m, 4H), 1.83 (tt, *J* = 6.5, 6.5 Hz, 2H), 1.59–1.38 (m, 4H), 0.19 (s, 9H); EI-MS (*m/z*) 318 (*M*⁺, 6), 203 (4), 183 (60), 169 (8), 107 (3), 91 (80), 77 (5), 73 (100).

2-(4-Benzoyloxybutyl)-1-(trimethylsilyloxy)cyclohexene 2k: 3.19 g, 96%; bp 155°C/0.03 mmHg; IR (neat) 2940, 2860, 1680, 1460, 1360, 1255, 1100, 950, 750 cm⁻¹; ¹H NMR (200 MHz, C₆D₆) δ 7.39–7.04 (m, 5H), 4.39 (s, 2H), 3.43 (t, *J* = 6.5 Hz, 2H), 2.25 (t, *J* = 6.4 Hz, 2H), 2.08–1.88 (m, 4H), 1.82–1.27 (m, 8H), 0.19 (s, 9H); EI-MS (*m/z*) 332 (*M*⁺, 10), 225 (2), 197 (2), 183 (53), 169 (9), 107 (3), 91 (76), 77 (7), 73 (100).

2-Hexyl-1-(trimethylsilyloxy)cycloheptene 2l: 2.14 g, 80%; bp 130°C/0.06 mmHg; IR (neat) 2940, 2860, 1685, 1450, 1345, 1255, 950, 750 cm⁻¹; ¹H NMR (200 MHz, C₆D₆) δ 2.44–2.34 (m, 2H), 2.31 (t, *J* = 7.3 Hz, 2H), 2.19–2.10 (m, 2H), 1.80–1.25 (m, 14H), 1.01 (t, *J* = 6.4 Hz, 3H), 0.28 (s, 9H); EI-MS (*m/z*) 268 (*M*⁺, 9), 253 (2), 225 (2), 197 (51), 183 (7), 73 (100).

Preparation of 5-butyl-1-(trimethylsilyloxy)bicyclo[3.1.0]hexane 3a. Representative Procedure. In a two necked flask equipped with a magnetic stirrer, were placed 2.12 g of silyl enol ether **2a** (10 mmol). After removal of air by a stream of argon, 11 mL of 1M solution of Et₂Zn (11 mmol) in hexane were then added at 25°C. After the white fumes had disappeared, 886 μL of diiodomethane (11 mmol) were added dropwise. An exothermic reaction occurred after a short induction period. This mixture was left stirring 15 min at 25°C then the two necked flask was equipped with a calcium chloride guard. A white precipitate appeared and after two hours the mixture was diluted with 20 mL of ethyl ether and ammonia was bubbled into the solution maintained at 0°C. Zinc salt-ammonia complexes were removed by filtration through Celite®. The filtrate was concentrated, and the product was purified by silica gel column chromatography (pentane/ethyl ether : 98/2) to give 1.69 g (75%) of silyloxybicyclohexane **3a**: IR (neat) 2940, 2860, 1450, 1350, 1250, 950, 750 cm⁻¹; ¹H NMR (200 MHz, CDCl₃) δ 2.06–1.81 (m, 2H), 1.69–1.15 (m, 10H), 1.03–0.75 (m, 3H), 0.60 (d, *J* = 5.3 Hz, 1H), 0.39 (dd, *J* =

5.3, 1.2 Hz, 1H), 0.14 (s, 9H); ^{13}C NMR (50.3 MHz, CDCl_3) δ 67.2, 34.5, 31.3, 31.1, 30.9, 30.4, 23.0, 20.1, 19.0, 14.2, 1.0; EI-MS (m/z) 226 (M^+ , 0.9), 211 (1), 183 (2), 169 (91), 73 (100).

($n+2$)-Substituted 1-(trimethylsilyloxy)bicyclo[$n.1.0$]alkanes **3b–3l** were prepared following the same procedure.

5-(3-Butenyl)-1-(trimethylsilyloxy)bicyclo[3.1.0]hexane 3b: 1.68 g, 75%; IR (neat) 2940, 2860, 1650, 1450, 1350, 1250, 950, 750 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 6.00–5.80 (m, 1H), 5.09–4.88 (m, 2H), 2.31–2.04 (m, 2H), 2.03–1.80 (m, 2H), 1.69–1.48 (m, 4H), 1.46–1.30 (m, 1H), 1.12–0.84 (m, 1H), 0.62 (dd, J = 5.5, 1.1 Hz, 1H), 0.41 (dd, J = 5.5, 1.5 Hz, 1H), 0.14 (s, 9H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 139.2, 113.7, 67.0, 34.4, 32.4, 31.4, 31.1, 30.4, 20.0, 19.0, 0.9; EI-MS (m/z) 244 (M^+ , 1.3), 183 (5), 169 (31), 73 (100).

5-(4-Chlorobutyl)-1-(trimethylsilyloxy)bicyclo[3.1.0]hexane 3c: 1.69 g, 65%; IR (neat) 2960, 2840, 1440, 1370, 1230, 950, 750 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 3.56 (t, J = 6.8 Hz, 2H), 2.04–1.75 (m, 4H), 1.62–1.42 (m, 6H), 1.42–1.20 (m, 2H), 0.62 (d, J = 5.4 Hz, 1H), 0.38 (d, J = 5.4 Hz, 1H), 0.14 (s, 9H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 139.2, 113.7, 67.0, 34.4, 32.4, 31.4, 31.1, 30.4, 20.0, 19.0, 0.9; EI-MS (m/z) 262 (M^+ , 0.3), 260 (M^+ , 0.8), 169 (72), 73 (100).

5-Hexyl-1-(trimethylsilyloxy)bicyclo[3.1.0]hexane 3d: 2.28 g, 90%; IR (neat) 2940, 2860, 1450, 1350, 1250, 950, 750 cm^{-1} ; ^1H NMR (200 MHz, CDCl_3) δ 2.04–1.80 (m, 2H), 1.67–1.12 (m, 14H), 0.89 (t, J = 6.8 Hz, 3H), 0.60 (d, J = 5.3 Hz, 1H), 0.39 (d, J = 5.3 Hz, 1H), 0.14 (s, 9H); ^{13}C NMR (50.3 MHz, CDCl_3) δ 66.8, 34.4, 33.6, 31.7, 30.9, 29.5, 29.1, 27.9, 23.5, 22.1, 19.7, 13.9, 0.8; EI-MS (m/z) 254 (M^+ , 6), 239 (4), 225 (2), 211 (2), 197 (2), 183 (21), 169 (12), 89 (2), 73 (100).

5-(3-Benzyloxypropyl)-1-(trimethylsilyloxy)bicyclo[3.1.0]hexane 3e: 2.51 g, 79%; IR (neat) 2940, 2860, 1460, 1360, 1260, 1100, 950, 750 cm^{-1} ; ^1H NMR (200 MHz, CDCl_3) δ 7.43–7.23 (m, 5H), 4.52 (s, 2H), 3.52 (t, J = 6.6 Hz, 2H), 2.06–0.90 (m, 10H), 0.61 (d, J = 5.4 Hz, 1H), 0.39 (dd, J = 5.4, 1.4 Hz, 1H), 0.14 (s, 9H); ^{13}C NMR (50.3 MHz, CDCl_3) δ 138.7, 128.2, 127.5, 127.3, 72.7, 70.6, 67.1, 34.4, 30.9, 30.4, 28.2, 28.0, 20.0, 18.9, 1.0; EI-MS (m/z) 229 (M^+ -OSiMe₃, 2), 183 (1), 169 (47), 107 (2), 91 (76), 77 (6), 73 (100).

5-(4-Benzyloxybutyl)-1-(trimethylsilyloxy)bicyclo[3.1.0]hexane 3f: 2.55 g, 77%; IR (neat) 2940, 2860, 1460, 1360, 1260, 1100, 950, 750 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 7.45–7.24 (m, 5H), 4.52 (s, 2H), 3.49 (t, J = 6.5 Hz, 2H), 2.05–1.77 (m, 2H), 1.77–1.15 (m, 8H), 1.10–0.80 (m, 2H), 0.61 (d, J = 5.5 Hz, 1H), 0.39 (dd, J = 5.5, 1.3 Hz, 1H), 0.14 (s, 9H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 138.7, 128.2, 127.5, 127.3, 72.8, 70.5, 67.1, 34.4, 31.3, 31.0, 30.7, 29.9, 24.7, 20.1, 19.0, 1.0; EI-MS (m/z) 332 (M^+ , 0.7), 241 (31), 183 (2), 169 (99), 107 (2), 91 (76), 77 (6), 73 (100).

5-(5-Benzyloxypentyl)-1-(trimethylsilyloxy)bicyclo[3.1.0]hexane 3g: 2.87 g, 83%; IR (neat) 2940, 2860, 1460, 1360, 1260, 1100, 950, 750 cm^{-1} ; ^1H NMR (200 MHz, CDCl_3) δ 7.44–7.23 (m, 5H), 4.54 (s, 2H), 3.52 (t, J = 6.4 Hz, 2H), 2.09–1.86 (m, 2H), 1.79–1.18 (m, 10H), 1.11–0.86 (m, 2H), 0.65 (d, J = 5.4 Hz, 1H), 0.39 (dd, J = 5.4, 0.7 Hz, 1H), 0.19 (s, 9H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 138.6, 128.2, 127.4, 127.3, 72.7, 70.4, 67.1, 34.4, 31.4, 30.9, 30.7, 29.7, 27.9, 26.4, 20.0, 18.9, 0.9.

6-Decyl-1-(trimethylsilyloxy)bicyclo[4.1.0]heptane 3i: 2.82 g, 87%; IR (neat) 2940, 2860, 1460, 1350, 1250, 950, 750 cm^{-1} ; ^1H NMR (200 MHz, CDCl_3) δ 2.16–1.97 (m, 2H), 1.97–0.96 (m, 24H), 0.88 (t, J = 6.4 Hz, 3H), 0.44 (d, J = 5.3 Hz, 1H), 0.39 (d, J = 5.3 Hz, 1H), 0.14 (s, 9H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 60.9, 34.9, 33.6, 31.9, 30.2, 30.1, 29.7, 29.6, 29.5, 29.4, 26.5, 25.3, 23.3, 22.7, 22.3, 22.1, 14.1, 1.3; EI-MS (m/z) 324 (M^+ , 4), 309 (2), 197 (7), 183 (100), 169 (4), 73 (65).

6-(3-Benzyloxypropyl)-1-(trimethylsilyloxy)bicyclo[4.1.0]heptane 3j: 2.99 g, 90%; IR (neat) 2940, 2860, 1455, 1360, 1250, 1100, 950, 750 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 7.47–7.22 (m, 5H), 4.54 (s, 2H), 3.52 (t, J = 6.5 Hz, 2H), 2.19–2.03 (m, 2H), 2.01–1.59 (m, 4H), 1.59–1.39 (m, 2H), 1.39–1.07 (m, 4H), 0.49 (d, J = 5.4 Hz, 1H), 0.45 (d, J = 5.4 Hz, 1H), 0.19 (s, 9H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 138.7, 128.1, 127.4, 127.2, 72.6, 70.7, 60.7, 33.3, 31.4, 29.9, 26.8, 24.6, 23.3, 22.2, 22.0, 1.2; EI-MS (m/z) 332 (M^+ , 16), 241

(5), 197 (17), 183 (10), 169 (16), 151 (42), 107 (11), 91 (100), 77 (4), 73 (73).

6-(4-Benzoyloxybutyl)-1-(trimethylsilyloxy)bicyclo[4.1.0]heptane 3k: 2.77 g, 80%; IR (neat) 2940, 2860, 1460, 1360, 1250, 1100, 950, 750 cm^{-1} ; ^1H NMR (200 MHz, CDCl_3) δ 7.45–7.22 (m, 5H), 4.52 (s, 2H), 3.49 (t, J = 6.5 Hz, 2H), 2.17–2.00 (m, 2H), 1.79–0.80 (m, 12H), 0.45 (d, J = 6.1 Hz, 1H), 0.41 (d, J = 6.1 Hz, 1H), 0.13 (s, 9H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 138.7, 128.3, 127.6, 127.4, 72.8, 70.6, 60.8, 34.7, 33.4, 30.1, 30.0, 25.0, 23.3, 23.1, 22.2, 22.0, 1.3; EI-MS (m/z) 346 (M^+ , 1.3), 197 (2), 183 (59), 107 (3), 91 (72), 89 (2), 77 (7), 73 (100).

7-Hexyl-1-(trimethylsilyloxy)bicyclo[5.1.0]octane 3l: 2.54 g, 90%; IR (neat) 2940, 2860, 1460, 1350, 1260, 950, 750 cm^{-1} ; ^1H NMR (200 MHz, CDCl_3) δ 2.36–2.13 (m, 1H), 2.13–1.92 (m, 1H), 1.92–1.04 (m, 18H), 0.89 (t, J = 6.5 Hz, 3H), 0.56 (d, J = 5.3 Hz, 1H), 0.41 (d, J = 5.3 Hz, 1H), 0.13 (s, 9H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 64.7, 39.4, 39.2, 34.3, 31.9, 31.8, 31.0, 29.8, 29.6, 28.8, 27.7, 26.2, 23.8, 14.0, 1.1; EI-MS (m/z) 282 (M^+ , 4), 267 (3), 253 (2), 239 (15), 225 (1), 211 (4), 197 (38), 73 (100).

Preparation of Racemic 5-butyl-1-(chloroacetoxy)bicyclo[3.1.0]hexane 5a. Representative Procedure. To a solution of 1.42 g of silyloxycyclopropane **3a** (6.3 mmol) in 10 mL of dichloromethane, maintained under argon, were added 6.3 mL of dry methanol and 8 μL of ClSiMe_3 (63 μmol). After 15 min at 25°C, TLC analysis showed that the reaction was over. After addition of 9 μL of triethylamine (63 μmol), the solvents were evaporated under reduced pressure. The flask containing the crude 5-butyl-1-bicyclo[3.1.0]hexanol **4a** was refilled with argon, and cooled at -30°C. Then a solution of 1.5 g of DMAP (12.3 mmol) in 9 mL of CH_2Cl_2 and a solution of 2.7 g of chloroacetic anhydride (15.8 mmol) in 9 mL of CH_2Cl_2 were added. The mixture was allowed to warm to 25°C, the reaction mixture becoming brown. After 2 h at 25°C, 5g of Celite® were added and CH_2Cl_2 was evaporated. The residue was taken up with 40 mL of ethyl ether and filtrated through Celite®. The filtrate was concentrated, and the product was purified by silica gel column chromatography (pentane/ether : 90/10) to give 1.06 g (73%) of chloroacetate **5a**: IR (neat) 2940, 2860, 1775, 1750, 1460 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 4.04 (s, 2H), 2.32–2.20 (m, 1H), 2.11–1.92 (m, 1H), 1.82–1.04 (m, 10H), 0.98 (d, J = 6.4 Hz, 1H), 0.91 (t, J = 6.8 Hz, 3H), 0.59 (d, J = 6.4 Hz, 1H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 167.5, 70.8, 40.9, 31.6, 31.5, 30.5, 30.3, 30.0, 22.7, 20.0, 18.7, 14.0; EI-MS (m/z) 230 (M^+ , 0.07), 95 (5), 93 (10), 79 (18), 77 (2), 51 (8), 49 (4), 41 (12).

Chloroacetates **5b–5l** were prepared following the same procedure.

5-(3-Butenyl)-1-(chloroacetoxy)bicyclo[3.1.0]hexane 5b: 1.08 g, 75%; IR (neat) 2940, 2860, 1775, 1750, 1650, 1460 cm^{-1} ; ^1H NMR (200 MHz, CDCl_3) δ 6.00–5.72 (m, 1H), 5.13–4.90 (m, 2H), 4.04 (s, 2H), 2.34–1.90 (m, 4H), 1.82–1.55 (m, 4H), 1.46–1.07 (m, 2H), 1.03 (d, J = 6.4 Hz, 1H), 0.62 (d, J = 6.4 Hz, 1H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 167.5, 138.7, 114.4, 70.7, 40.9, 32.1, 31.5, 31.4, 30.4, 30.3, 20.0, 18.5; EI-MS (m/z) 95 (33), 93 (70), 79 (22), 77 (51), 55 (82), 51 (16), 49 (26), 41 (100).

1-(Chloroacetoxy)-5-(4-chlorobutyl)bicyclo[3.1.0]hexane 5c: 1.52 g, 91%; IR (neat) 2940, 2860, 1775, 1750, 1460 cm^{-1} ; ^1H NMR (200 MHz, CDCl_3) δ 4.02 (s, 2H), 3.52 (t, J = 6.5 Hz, 2H), 2.32–2.11 (m, 1H), 2.10–1.97 (m, 1H), 1.97–1.42 (m, 7H), 1.41–1.02 (m, 3H), 0.97 (d, J = 6.3 Hz, 1H), 0.55 (d, J = 6.3 Hz, 1H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 167.5, 76.4, 70.6, 44.9, 40.8, 32.4, 31.2, 30.2, 29.6, 25.0, 20.0, 18.4.

1-(Chloroacetoxy)-5-hexylbicyclo[3.1.0]hexane 5d: 1.30 g, 80%; IR (neat) 2940, 2860, 1775, 1750, 1460 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 4.03 (s, 2H), 2.31–2.14 (m, 1H), 2.09–1.86 (m, 1H), 1.81–0.99 (m, 14H), 0.98 (d, J = 6.2 Hz, 1H), 0.88 (t, J = 6.1 Hz, 3H), 0.58 (dd, J = 6.2, 1.3 Hz, 1H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 167.6, 70.9, 40.9, 31.9, 31.8, 30.5, 30.4, 29.7, 29.4, 27.8, 22.6, 20.1, 18.5, 14.0; EI-MS (m/z) 95 (5), 93 (10), 79 (8), 77 (17), 51 (2), 49 (4); Anal. calcd for $\text{C}_{14}\text{H}_{23}\text{O}_2\text{Cl}$: C, 64.98; H, 8.96; Cl, 13.70, found: C, 65.15; H, 9.08; Cl, 13.61.

5-(3-Benzoyloxypropyl)-1-(chloroacetoxy)bicyclo[3.1.0]hexane 5e: 1.51 g, 72%; IR (neat) 2940, 2860, 1770, 1750, 1455, 1100 cm^{-1} ; ^1H NMR (200 MHz, CDCl_3) δ 7.39–7.24 (m, 5H), 4.51 (s, 2H), 4.02 (s, 2H),

3.50 (t, $J = 6.4$ Hz, 2H), 2.31–2.16 (m, 1H), 2.09–1.93 (m, 1H), 1.87–1.56 (m, 6H), 1.46–1.05 (m, 2H), 0.93 (d, $J = 6.3$ Hz, 1H), 0.62 (dd, $J = 6.3, 1.2$ Hz, 1H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 167.3, 138.3, 128.0, 127.3, 127.2, 72.4, 70.5, 69.9, 40.9, 31.0, 30.2, 30.1, 28.3, 26.7, 19.8, 18.1; EI-MS (m/z) 233 (2), 231 (6), 155 (28), 107 (3), 95 (5), 93 (9), 91 (100), 79 (7), 77 (13), 51 (2), 49 (3); Anal. calcd for $\text{C}_{18}\text{H}_{23}\text{O}_3\text{Cl}$: C, 66.97; H, 7.18; Cl, 10.98, found: C, 66.74; H, 7.23; Cl, 10.20.

5-(4-Benzoyloxybutyl)-1-(chloroacetoxy)bicyclo[3.1.0]hexane 5f: 1.74 g, 82%; IR (neat) 2940, 2860, 1775, 1750, 1460, 1100 cm^{-1} ; ^1H NMR (200 MHz, CDCl_3) δ 7.45–7.24 (m, 5H), 4.52 (s, 2H), 4.03 (s, 2H), 3.48 (t, $J = 6.3$ Hz, 2H), 2.25–2.15 (m, 1H), 2.15–1.85 (m, 1H), 1.85–1.04 (m, 10H), 0.99 (d, $J = 6.4$ Hz, 1H), 0.61 (dd, $J = 6.4, 1.4$ Hz, 1H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 167.3, 138.3, 128.0, 127.3, 127.1, 72.5, 70.4, 70.0, 40.6, 33.5, 31.3, 30.2, 30.1, 29.5, 24.3, 19.8, 18.1; EI-MS (m/z) 245 (5), 151 (9), 107 (5), 95 (3), 93 (7), 91 (100), 79 (10), 77 (30), 51 (1), 49 (4); Anal. calcd for $\text{C}_{19}\text{H}_{25}\text{O}_3\text{Cl}$: C, 67.75; H, 7.78; Cl, 10.22, found: C, 67.58; H, 7.31; Cl, 10.60.

5-(5-Benzoyloxypropyl)-1-(chloroacetoxy)bicyclo[3.1.0]hexane 5g: 1.61 g, 73%; IR (neat) 2940, 2860, 1775, 1750, 1460, 1100 cm^{-1} ; ^1H NMR (200 MHz, CDCl_3) δ 7.45–7.24 (m, 5H), 4.51 (s, 2H), 4.02 (s, 2H), 3.48 (t, $J = 6.4$ Hz, 2H), 2.35–2.12 (m, 1H), 2.12–1.88 (m, 1H), 1.88–1.05 (m, 12H), 0.98 (d, $J = 6.2$ Hz, 1H), 0.59 (d, $J = 6.2$ Hz, 1H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 167.4, 138.5, 128.1, 127.4, 127.3, 72.7, 70.6, 70.2, 40.8, 31.8, 30.5, 30.4, 30.2, 29.6, 27.5, 26.2, 19.9, 18.3; EI-MS (m/z) 261 (5), 259 (16), 183 (21), 107 (4), 95 (4), 93 (9), 91 (100), 79 (8), 77 (29), 51 (1), 49 (3); Anal. calcd for $\text{C}_{20}\text{H}_{27}\text{O}_3\text{Cl}$: C, 68.46; H, 7.76; Cl, 10.10, found: C, 69.25; H, 7.84; Cl, 9.93.

1-(Chloroacetoxy)-6-decylbicyclo[4.1.0]heptane 5i: 1.55 g, 75%; IR (neat) 2940, 2860, 1770, 1750, 1455 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 4.00 (s, 2H), 2.25–2.10 (m, 1H), 2.10–1.88 (m, 1H), 1.80–1.02 (m, 24H), 0.89 (t, $J = 6.6$ Hz, 3H), 0.70 (d, $J = 6.3$ Hz, 1H), 0.65 (d, $J = 6.3$ Hz, 1H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 167.2, 65.3, 41.0, 34.9, 31.8, 29.9, 29.8, 29.6, 29.3, 29.2, 27.5, 26.3, 25.9, 22.6, 21.9, 21.8, 21.2, 21.0, 14.1; EI-MS (m/z) 189 (1), 187 (2), 124 (3), 121 (22), 111 (100), 108 (80), 95 (15), 94 (9), 93 (35), 79 (11), 77 (25), 51 (1), 49 (2); Anal. calcd for $\text{C}_{19}\text{H}_{33}\text{O}_2\text{Cl}$: C, 69.61; H, 10.02; Cl, 10.69, found: C, 66.38; H, 10.11; Cl, 10.78.

6-(3-Benzoyloxypropyl)-1-(chloroacetoxy)bicyclo[4.1.0]heptane 5j: 1.93 g, 91%; IR (neat) 2940, 2860, 1770, 1750, 1450, 1100 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 7.38–7.22 (m, 5H), 4.51 (s, 2H), 3.96 (s, 2H), 3.49 (t, $J = 6.5$ Hz, 2H), 2.33–2.10 (m, 1H), 2.08–1.87 (m, 1H), 1.87–1.05 (m, 10H), 0.73 (d, $J = 6.8$ Hz, 1H), 0.68 (d, $J = 6.8$ Hz, 1H); ^{13}C NMR (50.3 MHz, CDCl_3) δ 167.3, 138.5, 128.3, 127.6, 127.5, 72.8, 70.3, 65.2, 41.0, 34.4, 29.7, 29.0, 26.6, 25.5, 21.9, 21.8, 21.1; EI-MS (m/z) 245 (3), 151 (21), 107 (4), 95 (4), 93 (9), 91 (100), 79 (10), 77 (18), 51 (2), 49 (5).

6-(4-Benzoyloxybutyl)-1-(chloroacetoxy)bicyclo[4.1.0]heptane 5k: 1.72 g, 78%; IR (neat) 2940, 2860, 1770, 1750, 1450, 1100 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 7.41–7.22 (m, 5H), 4.51 (s, 2H), 3.98 (s, 2H), 3.47 (t, $J = 6.3$ Hz, 2H), 2.28–2.09 (m, 1H), 2.05–1.83 (m, 1H), 1.75–1.05 (m, 12H), 0.72 (d, $J = 6.5$ Hz, 1H), 0.67 (d, $J = 6.5$ Hz, 1H); ^{13}C NMR (50.3 MHz, CDCl_3) δ 167.2, 138.6, 128.2, 127.5, 127.4, 72.8, 70.3, 65.2, 40.9, 34.7, 29.8, 29.7, 29.1, 25.8, 23.0, 22.0, 21.8, 21.1; EI-MS (m/z) 107 (4), 95 (7), 93 (16), 91 (100), 79 (8), 77 (20), 51 (2), 49 (4); Anal. calcd for $\text{C}_{20}\text{H}_{27}\text{O}_3\text{Cl}$: C, 68.46; H, 7.76; Cl, 10.10, found: C, 68.01; H, 7.85; Cl, 10.28.

1-(Chloroacetoxy)-7-hexylbicyclo[5.1.0]octane 5l: 1.44 g, 80%; IR (neat) 2940, 2860, 1770, 1750, 1460 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 4.02 (s, 2H), 2.59–2.42 (m, 1H), 2.21–2.03 (m, 1H), 1.90–1.06 (m, 18H), 1.00–0.85 (m, 4H), 0.77 (d, $J = 6.4$ Hz, 1H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 166.8, 69.3, 41.1, 34.6, 33.5, 31.7, 31.6, 31.5, 30.6, 20.4, 28.8, 27.1, 26.7, 25.8, 25.6, 13.9; EI-MS (m/z) 203 (0.7), 201 (1.5), 122 (21), 95 (13), 93 (30), 85 (3), 79 (8), 77 (19), 51 (7), 49 (18), 41 (100); Anal. calcd for $\text{C}_{16}\text{H}_{27}\text{O}_2\text{Cl}$: C, 66.01; H, 9.49; Cl, 13.34, found: C, 65.72; H, 9.47; Cl, 13.37.

Enzymatic Transesterification of 5-butyl-1-(chloroacetoxy)bicyclo[3.1.0]hexane 5a. Representative Procedure. In a flask equipped with a magnetic stirrer, were placed under argon 100 mg of the chloroacetate **5a** (0.43 mmol), and 500 mg of Lipozyme®. To the mixture were added 3 mL of *tert*-BuOMe and 886 μ L of dry 1-propanol (11.8 mmol) both previously purged of air by bubbling of argon. After 8h at 36°C, the reaction mixture was filtered under argon, and the solid was washed with air-free *tert*-BuOMe. After concentration under reduced pressure, the flask containing the residue was refilled with argon, 5 mL of ethyl ether were added and the mixture was cooled at -20°C. Then 243 μ L of triethylamine (1.7 mmol) and 221 μ L of chlorotrimethylsilane (1.7 mmol) were added. After 18h at 25°C, the reaction mixture was concentrated and the reaction products were separated by Florisil® column chromatography (pentane/ethyl ether : 90/10) to give 20 mg (42%) of silyloxycyclopropane **3a*** and 23 mg (47%) of chloroacetate **5a***.

Transesterifications of the chloroacetates **5b-5g** were made following the same procedure. Specific rotation of these optically active chloroacetates **5*** and silyloxybicycloalkanes **3*** are reported in Table 1.

Enzymatic Transesterification of 1-(chloroacetoxy)-6-decylbicyclo[4.1.0]heptane 5i. Representative Procedure. 52.6 mg of Chloroacetate **5i** (0.16 mmol) were dissolved in 0.7 mL of *tert*-BuOMe in a flask equipped with a magnetic stirrer. To this solution were added 210 mg of Lipozyme® and 330 μ L of dry 1-propanol (4.4 mmol). After 8h at 36°C, the reaction mixture was filtered and the solid was washed with *tert*-BuOMe. After concentration under reduced pressure, the reaction products were separated by column chromatography on neutral alumina activity III (pentane/ethyl ether : 90/10) to give 26 mg (49%) of chloroacetate **5i*** and 17.4 mg (43%) of 6-decylbicyclo[4.1.0]heptan-1-ol **4i***: IR (neat) 3420, 3060, 2930, 2860, 1460 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 2.22-2.07 (m, 1H), 1.79-1.68 (m, 2H), 1.60-1.02 (m, 24H), 0.88 (t, J = 6.6 Hz, 3H), 0.45 (s, 2H).

Transesterifications of the chloroacetates **5j-5l** were made following the same procedure. Specific rotation of these optically active chloroacetates **5*** and bicycloalkanols **4*** are reported in Table 1.

6-(3-Benzoyloxypropyl)bicyclo[4.1.0]heptan-1-ol 4j: IR (neat) 3400, 3090, 3060, 3030, 2940, 2860, 1460, 1100 cm^{-1} ; ^1H NMR (200 MHz, CDCl_3) δ 7.43-7.21 (m, 5H), 4.52 (s, 2H), 3.49 (t, J = 6.6 Hz, 2H), 2.24-2.09 (m, 1H), 1.85-1.04 (m, 12H), 0.47 (s, 2H).

6-(4-Benzoyloxybutyl)bicyclo[4.1.0]heptan-1-ol 4k: IR (neat) 3420, 3090, 3060, 3030, 2940, 2860, 1450, 1100 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 7.43-7.26 (m, 5H), 4.51 (s, 2H), 3.49 (t, J = 6.2 Hz, 2H), 2.23-2.10 (m, 1H), 1.98-1.08 (m, 14H), 0.47 (s, 2H).

7-Hexylbicyclo[5.1.0]octan-1-ol 4l: IR (neat) 3420, 3060, 2920, 2860, 1450 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 2.31 (dd, J = 6.6, 14.4 Hz, 1H), 1.95-1.78 (m, 1H), 1.78-1.10 (m, 19H), 0.89 (t, J = 6.8 Hz, 3H), 0.59 (d, J = 6.1 Hz, 1H), 0.46 (d, J = 6.1 Hz, 1H).

Rearrangement of chloroacetate 5a* and silyloxybicyclohexane 3* into 2-butyl-2-methylcyclopentanone 6a. Representative Procedure. 23 mg of optically active chloroacetate **5a*** (0.1mmol) were dissolved in 200 μ L of methanol and 100 μ L of 2N NaOH solution (0.2 mmol) were added. This mixture was stirred 4h at 60°C. The product was extracted with ethyl ether, and the organic solution was washed with brine and dried (Na_2SO_4). After concentration the crude product was purified by column chromatography on silica gel (pentane/ethyl ether : 90/10) to give 12.3 mg (80%) of 2-butyl-2-methylcyclopentanone (-)-**6a**: IR (neat) 2960, 2840, 1750, 1460 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 2.42-2.13 (m, 2H), 2.02-1.64 (m, 4H), 1.52-1.09 (m, 6H), 0.99 (s, 3H), 0.89 (t, J = 6.9 Hz, 3H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 222.9, 47.7, 36.4, 35.6, 29.7, 26.5, 23.3, 21.8, 18.7, 14.0; EI-MS (m/z) 154 (M^+ , 29), 111 (17), 98 (100), 97 (11), 55 (55), 43 (25), 41 (41); Anal. calcd for $\text{C}_{10}\text{H}_{18}\text{O}$: C, 77.87; H, 11.76, found: C, 78.18; H, 11.73; $[\alpha]_{\text{D}}^{20}$ = -39° (c = 1.9, THF, ee = 97%). Rearrangement of silyloxycyclopropane **3a*** made using the same procedure gave mainly the enantiomer (+)-**6a** (yield 85%): $[\alpha]_{\text{D}}^{20}$ = +47° (c = 0.8, THF, ee = 98%). Measurements of the ee's of **6a** were made by GC: 80°C,

0.75 bar: (-)-(R), T_R = 23.32 min; (+)-(S), T_R = 26.51 min.

Rearrangements of **6b***-**6l***, **3b***-**3g*** and **4h***-**4l*** were made following the same procedure.

2-(3-Butenyl)-2-methylcyclopentanone 6b: IR (neat) 2960, 2840, 1750, 1650, 1460 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 5.89–5.67 (m, 1H), 5.09–4.88 (m, 2H), 2.63–2.41 (m, 2H), 2.19–1.64 (m, 6H), 1.52–1.48 (m, 2H), 1.02 (s, 3H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 223.4, 128.5, 114.5, 48.1, 37.7, 35.7, 35.6, 28.6, 21.7, 18.7; Anal. calcd for $\text{C}_{10}\text{H}_{16}\text{O}$: C, 78.90; H, 10.59; O, 10.51, found: C, 79.11; H, 10.51; O, 10.47. From **5b*** was obtained (-)-**6b** (Yield 85%): $[\alpha]_D^{20}$ = -36.8° (c = 0.8, THF, ee = 97%) and from **3b*** the compound (+)-**6b** (Yield 92%): $[\alpha]_D^{20}$ = $+32.5^\circ$ (c = 0.8, THF ee = 86%). The 2-(3-butenyl)-2-methylcyclopentanone **6b** was transformed by hydrogenation (1 bar, Pd-C 10%, EtOH) into 2-butyl-2-methylcyclopentanone **6a** for the measurement of the ee's (vide supra).

2-(4-Chlorobutyl)-2-methylcyclopentanone 6c: IR (neat) 2960, 2840, 1750, 1460 cm^{-1} ; ^1H NMR (250 MHz, CDCl_3) δ 3.54 (t, J = 6.7 Hz, 2H), 2.49–2.12 (m, 2H), 1.99–1.64 (m, 5H), 1.61–1.22 (m, 5H), 1.01 (s, 3H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 228.4, 48.2, 44.8, 37.6, 35.8, 35.5, 32.9, 21.8, 21.7, 18.7; Anal. calcd for $\text{C}_{10}\text{H}_{17}\text{ClO}$: C, 63.65; H, 9.08; Cl, 18.79; O, 8.48, found: C, 63.82; H, 9.15; Cl, 18.78; O, 8.61. From **5c*** was obtained (-)-**6c** (Yield 88%): $[\alpha]_D^{20}$ = -36.1° (c = 1.2, THF, ee = 93%) and from **3c*** the compound (+)-**6c** (Yield 90%): $[\alpha]_D^{20}$ = $+33.6^\circ$ (c = 0.5, THF, ee = 94%). Measurements of the ee's of **6c** were made by GC: 100°C, 0.8 bar: (-)-(S), T_R = 93.7 min; (+)-(R), T_R = 126.9 min.

2-Hexyl-2-methylcyclopentanone 6d: IR (neat) 2960, 2840, 1750, 1460 cm^{-1} ; ^1H NMR (200 MHz, CDCl_3) δ 2.40–2.10 (m, 2H), 2.00–1.61 (m, 4H), 1.48–1.07 (m, 10H), 0.98 (s, 3H), 0.87 (t, J = 6.7 Hz, 3H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 224.6, 48.3, 37.7, 36.7, 35.6, 31.7, 29.8, 24.2, 22.6, 21.8, 18.7, 14.1; EI-MS (m/z) 182 (M^+ , 1.9), 98 (100), 97 (11), 71 (3), 57 (11), 55 (29); Anal. calcd for $\text{C}_{12}\text{H}_{22}\text{O}$: C, 79.06; H, 12.16; O, 8.78, found: C, 79.27; H, 12.19; O, 8.81. From **5d*** was obtained (-)-**6d** (Yield 86%): $[\alpha]_D^{20}$ = -26.0° (c = 0.5, THF, ee = 81%) and from **3d*** the compound (+)-**6d** (Yield 90%): $[\alpha]_D^{20}$ = $+32.7^\circ$ (c = 1, THF ee = 97%). Measurements of the ee's of **6d** were made by GC: 90°C, 0.75 bar: (-)-(R), T_R = 45.8 min; (+)-(S), T_R = 49.5 min.

2-(3-Benzoyloxypropyl)-2-methylcyclopentanone 6e: IR (neat) 2940, 2860, 1740, 1460, 1100 cm^{-1} ; ^1H NMR (200 MHz, CDCl_3) δ 7.42–7.14 (m, 5H), 4.49 (s, 2H), 3.46 (t, J = 6.7 Hz, 2H), 2.40–2.12 (m, 2H), 1.98–1.37 (m, 8H), 1.01 (s, 3H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 222.6, 128.1, 127.8, 127.1, 127.0, 72.3, 70.1, 47.4, 37.1, 35.2, 32.6, 24.3, 21.3, 18.2; EI-MS (m/z) 139 (7), 111 (1), 107 (7), 98 (35), 97 (11), 91 (100), 77 (6), 55 (14); Anal. calcd for $\text{C}_{16}\text{H}_{22}\text{O}_2$: C, 78.01; H, 9.00, found: C, 78.17; H, 8.92. From **5e*** was obtained (-)-**6e** (Yield 87%): $[\alpha]_D^{20}$ = -29.8° (c = 1, THF, ee = 91%) and from **3e*** the compound (+)-**6e** (Yield 92%): $[\alpha]_D^{20}$ = $+24.8^\circ$ (c = 1.7, THF ee = 87%). Measurements of the ee's of **6e** were made by ^1H NMR in the presence of $\text{Eu}(\text{hfc})_3$: the methyl singulets (δ 1.01 ppm) were shifted to 2.56 [(-)-(S)] and 2.58 ppm [(+)-(R)].

2-(4-Benzoyloxybutyl)-2-methylcyclopentanone 6f: IR (neat) 2940, 2860, 1740, 1460, 1105 cm^{-1} ; ^1H NMR (200 MHz, CDCl_3) δ 7.42–7.20 (m, 5H), 4.50 (s, 2H), 3.46 (t, J = 6.4 Hz, 2H), 2.39–2.11 (m, 2H), 1.99–1.15 (m, 10H), 0.99 (s, 3H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 223.5, 138.4, 128.2, 127.5, 127.3, 72.7, 69.9, 48.1, 37.5, 36.3, 35.4, 30.1, 21.6, 20.8, 18.5; EI-MS (m/z) 260 (M^+ , 0.8), 153 (2), 111 (6), 107(9), 98 (38), 97 (9), 91 (100), 77 (6), 55 (15); Anal. calcd for $\text{C}_{17}\text{H}_{24}\text{O}_2$: C, 78.42; H, 9.29, found: C, 78.11; H, 9.11. From **5f*** was obtained (-)-**6f** (Yield 85%): $[\alpha]_D^{20}$ = -37.0° (c = 1, THF, ee = 95%) and from **3f*** the compound (+)-**6f** (Yield 92%): $[\alpha]_D^{20}$ = $+28.3^\circ$ (c = 1, THF, ee = 85%). Measurements of the ee's of **6f** were made by GC: 160°C, 0.75 bar: (-)-(R), T_R = 202.3 min; (+)-(S), T_R = 206.7 min.

2-(5-Benzoylopentyl)-2-methylcyclopentanone 6g: IR (neat) 2940, 2860, 1740, 1460, 1105 cm^{-1} ; ^1H NMR (200 MHz, CDCl_3) δ 7.40–7.19 (m, 5H), 4.48 (s, 2H), 3.46 (t, J = 6.4 Hz, 2H), 2.38–2.05 (m, 2H), 1.98–1.07 (m, 12H), 0.98 (s, 3H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 223.0, 138.3, 127.9, 127.2, 127.0, 72.4, 69.9, 47.8, 37.2, 36.2, 35.2, 29.3, 26.4, 23.7, 21.4, 18.3; EI-MS (m/z) 274 (M^+ , 2), 153 (2), 111 (16), 107 (15), 98 (90), 97 (11), 91 (100), 77 (4), 55 (11); Anal. calcd for $\text{C}_{18}\text{H}_{26}\text{O}_2$: C, 78.83; H, 8.76, found: C, 79.08; H, 8.48.

From **5g*** was obtained (-)-**6g** (Yield 86%): $[\alpha]_D^{20} = -25.2^\circ$ ($c = 1.6$, THF, ee = 90%) and from **3g*** the compound (+)-**6g** (Yield 91%): $[\alpha]_D^{20} = +22.6^\circ$ ($c = 1$, THF, ee = 86%). Measurements of the ee's of **6g** were made by ^1H NMR in the presence of $\text{Eu}(\text{hfc})_3$: the methyl singulets (δ 0.98 ppm) were shifted to 2.60 [(-)-(**R**)] and 2.68 ppm [(+)-(**S**)].

2-Decyl-2-methylcyclohexanone **6i**: IR (neat) 2940, 2860, 1715, 1455 cm^{-1} ; ^1H NMR (200 MHz, CDCl_3) δ 2.49–2.25 (m, 2H), 2.02–1.59 (m, 6H), 1.49–1.16 (m, 18H), 1.12 (s, 3H), 0.89 (t, $J = 6.7$ Hz, 3H); ^{13}C NMR (50.3 MHz, CDCl_3) δ 216.3, 48.6, 39.5, 38.8, 37.5, 31.9, 30.3, 29.6, 29.5, 29.4, 29.3, 27.5, 23.7, 22.7, 22.5, 21.0, 14.1; EI-MS (m/z) 252 (M^+ , 0.3), 113 (36), 112 (100), 111 (7), 85 (3), 71 (4), 69 (23), 57 (14), 55 (12), 43 (43); HRMS, calcd for $\text{C}_{17}\text{H}_{32}\text{O}$: 252.2453, found: 252.2456. From **5i*** was obtained (-)-**6i** (Yield 85%): $[\alpha]_D^{20} = -31.0^\circ$ ($c = 0.7$, THF, ee = 74%) and from **4i*** the compound (+)-**6i** (Yield 90%): $[\alpha]_D^{20} = +35.0^\circ$ ($c = 0.6$, THF, ee = 85%). Measurements of the ee's of **6i** were made by GC: 130°C , 0.75 bar: (-)-(**R**), $T_R = 220.6$ min; (+)-(**S**), $T_R = 227.5$ min.

2-(3-Benzoyloxypropyl)-2-methylcyclohexanone **6j**: IR (neat) 2940, 2860, 1715, 1460, 1100 cm^{-1} ; ^1H NMR (200 MHz, CDCl_3) δ 7.42–7.20 (m, 5H), 4.50 (s, 2H), 3.46 (t, $J = 6.0$ Hz, 2H), 2.53–2.39 (m, 2H), 2.04–1.25 (m, 10H), 1.06 (s, 3H); ^{13}C NMR (62.9 MHz, CDCl_3) δ 215.9, 138.4, 127.5, 128.3, 127.4, 72.7, 70.5, 48.2, 39.2, 38.6, 33.9, 27.4, 24.1, 22.4, 20.9; EI-MS (m/z) 260 (M^+ , 0.4), 153 (17), 112 (34), 111 (6), 107 (6), 91 (100), 77 (4), 55 (12); Anal. calcd for $\text{C}_{17}\text{H}_{24}\text{O}_2$: C, 78.46; H, 9.23, found: C, 78.25; H, 8.97. From **5j*** was obtained (-)-**6j** (Yield 88%): $[\alpha]_D^{20} = -38.6^\circ$ ($c = 1$, THF, ee = 90%) and from **4j*** the compound (+)-**6j** (Yield 90%): $[\alpha]_D^{20} = +36.3^\circ$ ($c = 1$, THF, ee = 86%). Measurements of the ee's of **6j** were made by ^1H NMR in the presence of $\text{Eu}(\text{hfc})_3$: the methyl singulets (δ 1.06 ppm) were shifted to 2.63 [(-)-(**S**)] and 2.65 ppm [(+)-(**R**)].

2-(4-Benzoyloxybutyl)-2-methylcyclohexanone **6k**: IR (neat) 2940, 2860, 1715, 1450, 1100 cm^{-1} ; ^1H NMR (200 MHz, CDCl_3) δ 7.32–7.15 (m, 5H), 4.51 (s, 2H), 3.46 (t, $J = 6.3$ Hz, 2H), 2.21–2.07 (m, 2H), 1.68–1.06 (m, 12H), 1.01 (s, 3H); ^{13}C NMR (50.3 MHz, CDCl_3) δ 215.9, 138.5, 128.2, 127.6, 127.4, 72.8, 70.1, 48.5, 39.2, 38.7, 37.3, 30.3, 27.4, 22.5, 20.9, 20.4; EI-MS (m/z) 274 (M^+ , 1.1), 167 (2.1), 112 (100), 111 (8), 107 (6), 91 (95), 77 (4), 55 (55); Anal. calcd for $\text{C}_{18}\text{H}_{26}\text{O}_2$: C, 78.79; H, 9.55, found: C, 79.02; H, 9.75. From **5k*** was obtained (-)-**6k** (Yield 88%): $[\alpha]_D^{20} = -31.0^\circ$ ($c = 1.4$, THF, ee = 85%) and from **4k*** the compound (+)-**6k** (Yield 91%): $[\alpha]_D^{20} = +24.0^\circ$ ($c = 0.9$, THF, ee = 70%). Measurements of the ee's of **6k** were made by GC: 165°C , 0.75 bar: (-)-(**S**), $T_R = 199.2$ min; (+)-(**R**), $T_R = 201.9$ min.

2-Hexyl-2-methylcycloheptanone **6l**: IR (neat) 2940, 2860, 1710, 1455 cm^{-1} ; ^1H NMR (200 MHz, CDCl_3) δ 2.79–2.49 (m, 1H), 2.42–2.25 (m, 1H), 1.92–1.13 (m, 18H), 1.03 (s, 3H), 0.87 (t, $J = 6.2$ Hz, 3H); ^{13}C NMR (50.3 MHz, CDCl_3) δ 217.9, 66.7, 50.9, 40.2, 37.7, 30.7, 29.9, 28.4, 26.6, 24.4, 23.9, 22.5, 21.4, 14.0; EI-MS (m/z) 139 (1.3), 126 (100), 125 (3), 111 (8), 97 (27), 83 (16), 69 (44), 57 (23), 55 (83), 43 (23); Anal. calcd for $\text{C}_{14}\text{H}_{26}\text{O}$: C, 79.94; H, 12.46; O, 7.61, found: C, 79.41; H, 12.21; O, 7.86. From **5l*** was obtained (-)-**6l** (Yield 82%): $[\alpha]_D^{20} = -20.7^\circ$ ($c = 2.8$, THF, ee = 51%) and from **4l*** the compound (+)-**6l** (Yield 86%): $[\alpha]_D^{20} = +33.0^\circ$ ($c = 0.8$, THF, ee = 87%). Measurements of the ee's of **6l** were made by GC: 100°C , 0.75 bar: (-)-(**R**), $T_R = 111.5$ min; (+)-(**S**), $T_R = 116.7$ min.

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- For a review on "Asymmetric Creation of Quaternary Carbon Centers" which presents most part of the methods to prepare optically active cycloalkanones, see: Fuji, K. *Chem. Rev.* **1993**, *93*, 2037–2066.

References for other methods, other examples or more recent results are indicated for each item.

5. Some specific methods allow the preparation of enantiomerically enriched α,α -disubstituted cyclobutanones: the rearrangement of optically active oxaspiropentanes,⁴ diastereoselective [2+2] cycloadditions⁴ and rearrangement of optically active cyclopropylmethyl alcohol derivatives⁴ (see also: Nemoto, H.; Yamada, T.; Ishibashi, H.; Takazawa, J.; Fukumoto, K. *Heterocycles* **1991**, 32, 863-866).
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25. Handling of the bicycloalkanols was made under argon. This precaution is particularly important in the case of bicyclo[3.1.0]hexan-1-ols which are air sensitive.²⁹
26. Careful neutralisation of the reaction mixture before evaporation of methanol gave a more reliable method to obtain the bicycloalkanol free of α -methylketone resulting from cyclopropanic bond cleavage which is easier in acid or basic medium.
27. These chloroacetates were mixed with regioisomers which came from the minor less substituted silyl enol ether formed during the synthesis of **2**.
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31. The transesterification of **5I** was also attempted in the presence of other lipases. With the lipases from *Candida lipolytica*, from *Pseudomonas fluorescens* and from *Chromobacterium viscosum* no reaction with 1-propanol were observed after 3 days at 36°C (these results confirm that the uncatalyzed transesterification was not a competitive process in our conditions). With the lipase from *Candida rugosa*, the lipase from *Pseudomonas cepacia* and the pig pancreatic lipase the reaction has occurred and the conversions were respectively 7% (after 3 days), 13% (after 3 days) and 24% (after 4 days) using the same ratio of the enzyme weight to the chloroacetate weight than that of the Lipozyme®-catalyzed reaction.
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